

# Gene Responsible for Emergence of Antifungal Resistance in *Candida* Spps. & Diagnosis: A Review

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## Abstract

*Candida* is a commensal & present in moist region of the human body. It also acts as an opportunistic pathogen that is prevalent in immunocompromised patients such as in case of HIV, hepatitis B, Cancer or also in diabetic patients. In COVID19 era, it also causes no. of cases in COVID19 patients due to immunosuppression. It is also known by name i.e. white fungus. The numbers of cases mainly increase in patients who are hospitalized due to the COVID19, because in hospital there are number of resistance organisms which are present in hospital environment. *C. auris* is reported in India but also in all over the world. Rise of antifungal resistance in *Candida* also become a big problem for public health

sector. Due to this, *Candida* acquired resistance against azole, polyenes & echinocandins. This resistance mechanism increased due to over expression of genes such as SNQ2, TPO3, ABC1, ERG11, ERG2, ERG3, ERG5, ERG6 which gain in function through mutation in the transcription that leads to alteration in gene expression that cause increase in drug efflux pumps, increased concentration of lanosterol 14 $\alpha$ -demethylase & point alteration in which ergosterol content decreased in cells. Diagnosis can be done by multiplex PCR technique, to know the gene which responsible for antifungal drug resistance in that particular *Candida* specie.

**Keywords:** Candidemia; Antifungal drugs; COVID19; Co-infection; CHROMagar.

## Introduction

Fungal infection are commonly neglected in the presence of bacteria & viruses but there are 1.5 million fungal species in which 300 causes human diseases & 300 million people are infected by these invasive fungal pathogens & 1.6 million die annually due to these pathogens including *Aspergillus*, *Nocardia*, *Mucormycosis*, *Cryptococcus*, *Basidiomycosis*, *Pneumocystis*, *Candida* & many more.<sup>1</sup> *Candida* is a Dimorphic Fungi & also called as Yeast like fungi, which present as a commensal in human body at moist region i.e. gastrointestinal, genitourinary tracts and in the oral and conjunctival flora.<sup>2</sup>

*Candida* genera contain mainly 200 species in which the medically important species includes *Candida albicans*, *Candida glabrata*, *Candida tropicalis*, *Candida parapsilosis*, and *Candida krusei* & all these are responsible for 90% invasive infections in humans but there are some species which also emerge as pathogens such as *Candida guilliermondii*, *Candida kefyr*, *Candida rugosa*, *Candida dubliniensis*, and *Candida famata*.

It has three main cellular morphologies: yeast, pseudohyphae, and hyphae. Yeast are single cells that are oval in shape and divided by budding when grown at room temperature. Pseudohyphae and Hyphae are filamentous form which grown in polarized manner in host body or at 37°C. This form

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also provide it another virulence factor because in hyphal form are able to release hydrolytic enzyme such as phospholipid & proteinase which help to invade the organism to adjacent tissue specifically in endothelial & epithelial cells.<sup>3,7</sup>

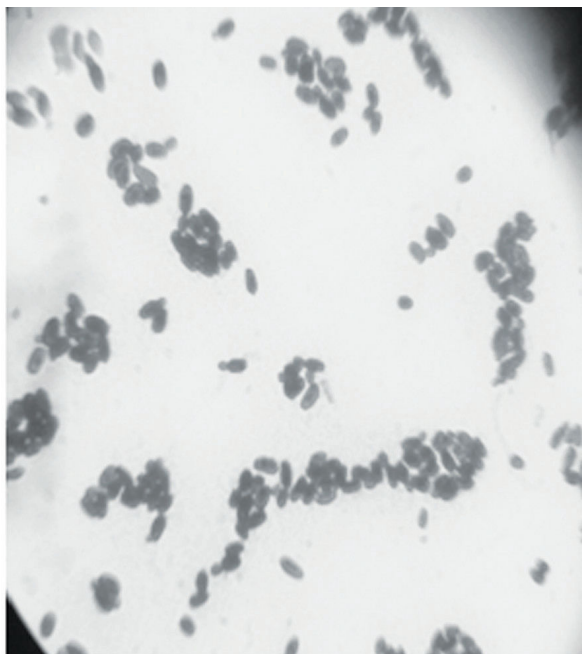


Fig. 1: microscopic view of *Candida* under light microscope.

*Candida* a commensal not allows the other organisms to settle down in body which act as defense for invading organisms but in an immunosuppressive conditions such as in cases of HIV, Hepatitis, Cancer, Diabetic condition, Chemotherapy, Organtransplantation & even in era of COVID19, it act as an invasive organism which causes co-infection in patients mostly to those who area dmittedtohospitals.

*Candida* causes 3 type of infection such as superficial, Subcutaneous & invasive infection with a mortality rate of 45%, according to Nosocomial Infections Surveillance System (NNISS) reported that *Candida* cause four the number of blood Infection known as Candidemia which is life threatening Infection.<sup>4</sup>

Superficial Candidiasis are most common in several countries but, its cases are high in tropical & sub tropical countries. This includes commonly infections of mucosal layer of human body such as nail, vagina, oralthrush, paronychia & inter digital candidiasis on hand or foot.<sup>5</sup>

Subcutaneous candidiasis infection of humans results from inoculation of *Candida* in tissues which results from trauma, injury or hematogenous spread & these infections also responsible for invasive infection.<sup>6</sup>

Invasive candidiasis infection increased significantly in Immune compromised population who are hospitalized due to Nosocomial Infection.

**Table 1:** Most prevalent type of infections of *Candida* in humans.

#### **Candida Infection Rate**

Rank	Type of Infection
3-4th	Most isolated nosocomial blood stream infection.
4th	Most common Hospital Acquired Systemic Infection.
5th	Most common cause of blood stream infections in pediatric intensive care units.

It's risk factors includes hematological malignances, bone marrow transplantation, prolong treatment with corticosteroid, in Intensive care, chemotherapy, HIV infection or incase of malnutrition & in severe burn.<sup>7</sup>

Invasive candidias is an important health issue that caused by several *Candidaspps*. Commonly *C.albicans* i.e. 70 percent but varies according to geographical condition. Through bloodstream it spread to other organs such as the liver, heart, kidney, spleen & brain.<sup>8</sup>

Mainly the infection of *Candida* to human are prevalent in hospitalized people who already surffed from another disease & in present scenario it also seen in many patients. When human immune suppressed due to this disease then self & environment organism try to override the human body that leads to Co-infection & with this hospital environment may also contains antibiotics Resistance organisms which cause disease in human. And anti fungal resistance *Candida* also responsible for infection in this kind of situation, through Horizontal Gene Transfer it also transfer gene which spread resistance to other organisms & make the patient a Carrier who can also responsible for the spread of community acquire infection when it was discharged from hospital.

There are many antifungals used to treat *Candida* infection & mainly are azoles antifungals which are easily available, inexpensive & exhibits limited toxicity but due to over & unwanted use of these drugs increased the emergence of antifungal resistance *Candida*. Azoles resistance infection caused by *Candida albicans* and but this resistance also emerge in non albicansspps.<sup>9</sup>

There are also several classes of compounds used for treatment of *Candida* infections which explains in below table.<sup>10-17</sup>

**Table 2:** Various classes of antifungal with their different properties.

Classes	Drugs	Dosage		Mechanism of effect	Mechanism of resistance	Toxicity
		IV	Oral			
Azoles	Fluconazole	400–800 mg/d 100–200 mg/dc	400–800 mg/d 100–200 mg/dc	impair ergo sterol synthesis by inhibiting C14-a sterol demethylase that lead to disruption of sterol precursors & reduction of registered in cell membrane.	a) Alteration in ERG11 & THR 1 gene cause modification in quality & quantity of 14a-demethylase in the expression of resistance to azole antifungal agents. b)the up regulation of efflux pumps, has also promote drug resistance via a decrease in intracellular drug levels.	the most common side-effects include rash, headache, or gastrointestinal upset & Hepatotoxicity.
	Itraconazole	NA	200 mg 1-3/d	-	-	-
	Voriconazole	6 mg/kg for 2 doses, then 4 mg/kg q 12 h	400 mg bid for 2 doses, then 200 mg q 12 h	-	-	-
Polynes	Amphotericin B	0.7–1 mg/kg/da	NA	Interaction with membrane sterols results in production of pore that leads to altered permeability & leakage cause death of organisms.	Defects in the ERG3 gene involved in ergosterol biosynthesis lead to accumulation of other sterols in the fungal membrane.	Adverse effects include renal toxicity, infusion reactions, electrolyte abnormalities, and hepatotoxicity.
Allylamines	Naftifine	NA	250 mg day or topical (1% cream) administration	inhibit ergo sterol synthesis at the level of squaleneepoxidase with highly selective for the fungal enzyme but minimal effect on mammalian cholesterol synthesis.	Not yet reported	Patients experiencing AE, including mild burning/stinging, itching, erythema, irritation, and rarely, allergic reactions.
	Terbinafine	NA	7–12.5 mg/kg/day	-	-	-
Echino-candins	Caspofungin	70 mg for 1 dose, then 50 mg/d	NA	compounds disrupt the fungal cell wall by inhibiting the synthesis of b-1,3 glucan which a fungal cell polysaccharide.	Hot-spot mutations in FKS1 or FKS2 genes cause change in amino acids substitution of fks subunits of glucansynthase.	adverse reactions include gastrointestinal upset, headache, elevation of liver (aminotransferase) tests, or mild infusion reaction.
	Micafungin	100–150 mg/d 50 mg/d	NA	-	-	-
	Anidulafungin	100–200 mg for 1 dose, then 50–200 mg/d	NA	-	-	-

Antifungal activity can be measure with standard dilution in liquid media or with solid surface agar with drug gradient can be used. Two major protocols are currently used i.e CLSI (clinical laboratory standard institute) & EUCAST (European committee on antimicrobial susceptibility testing), there protocols yields called MIC (Minimum inhibitor concentration).<sup>18</sup> MIC is the lowest concentration (mg/l) of antibiotics

which is able to inhibit the growth of organism & also helpful to know the resistance or susceptibility of organism for against a one or more antibiotics.

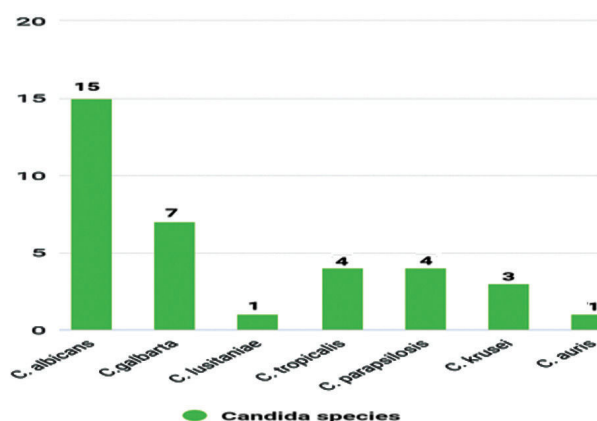
MIC play important role in new antifungal drugs which used in vitro testing that can predict the effects of compounds in vivo & further clinical results used for treatment.<sup>19</sup>

Mechanisms leading to the emergence of

resistance includes single-point mutations chromosomal rearrangements & horizontal gene transfer (HGT) or hybridization but in all these HGT is most responsible for transfer of gene which are associated with resistance emergence.<sup>20</sup>

**Table 3:** Different type of gene found in different - different candida species which are responsible for emergence of antifungal resistance.

Antifungal Classes	Species	Gene	
Azoles	<i>C. albicans</i>	ERG11	
		UPC2	
		TAC1	
		MRR1	
		ERG3	
		CDR1	
		CDR2	
		SNQ2	
		ABC1	
		MDR1	
		TPO3	
		<i>C.glabrata</i>	MDR1
			TPO3
		<i>C.parapsilosis</i>	ERG11
			CDR1
CDR2			
SNQ2			
ABC1			
MDR1			
<i>C.tropicalis</i>	ERG11		
	CDR1		
	CDR2		
	SNQ2		
<i>C.kruse</i>	ABC1		
	MDR1		
	TPO3		
	ERG11		
	CDR1		
<i>C.auris</i>	CDR2		
	SNQ2		
	ABC1		
	MDR1		
	TPO3		
Echino-candins	<i>C. albicans</i>	FKS1	
		FKS2	
Polyenes	<i>C. albicans</i>	ERG2	
		ERG3	
		ERG5	
		ERG11	
		<i>C. glabrata</i>	ERG2
			ERG6



**Fig. 2:** Graphical representation of no. of gene present in different Candida species.

Co-infection of Candida is seen in COVID 19 patients since the pandemic started & co-infection also named as super infection. *C. glabrata* a common fungal commensal of mucosal surface that cause blood stream infection in some countries includes USA, Asia & European countries with multidrug resistance that shows the high tolerance of Candida spp. Against different classes of antifungal drugs.<sup>21</sup> SARS-CoV2 is responsible for pandemic all over the world which effect approximately 196 countries people & started as epidemic in Wuhan, China. Wuhan virus isolated from epithelial cells of nasopharynx from a cluster of patient with pneumonia like symptoms. SARS-CoV2 belong to corona virus family but it is different from both MERS-CoV & SARS-CoV, these also infect humans. It emerges as global pathogen which challenges the whole world public health sector. Before SARS-CoV2 pandemic start, SARS-CoV also outbreaks in 2002 & 2003 in changing province china while MERS-CoV was outbreaks in 2012 in middle East. In late Dec 2019, SARS-CoV2 outbreaks all over the world which lead to biggest pandemic of the century after the Spanish flu 1920 which responsible for death of 50 million people in all over the world.<sup>22</sup>

During April-July 2020, 15 patients who was critically ill due to COVID19 they are affected with candidemia which was caused by *C.auris* with 60 percent high case fatality rate.<sup>23</sup> COVID19 patients are prone to respiratory disease syndrome which are acquired by patients through ventilators in ICUs & this also reported in 40 countries across 6 continents that's why it also known as environmental colonizer of ICUs but diagnostics resources are limited in another countries.<sup>24</sup>

Primary antifungals are failed against the multidrug resistance candidaspps. Such as *C.auris* & *C.glabrata* which increase the demand for

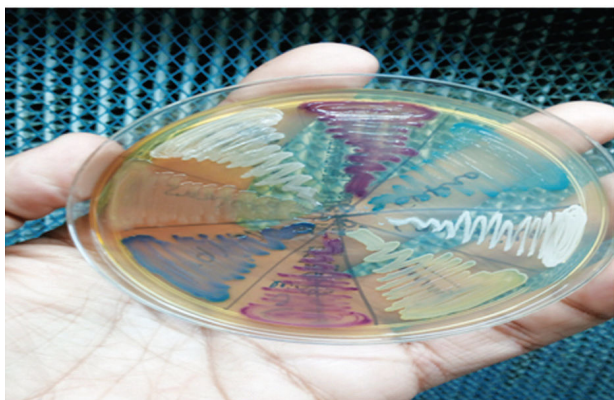


development of new antifungals with different novel action mechanism against organisms.<sup>25</sup>

Diagnosis of *Candida* spp. infection done by conventional detection method that based on blood & culture plate method which is time consuming and take 2-4 days to identify *Candida* spp. While nonconventional method include serological & nucleic acid detective test. For diagnosis specimen collected from site of infection such as nail clippings, skin scrapings, tissues, aspirates and respiratory specimens, microscopic observation and detection can be used, whereas for fluidic samples such as blood and urine.<sup>26</sup> For isolation of *Candida*, specimen is cultured general in Sabouraud dextrose agar (SDA) & Potato dextrose agar (PDA) but for advance diagnosis today CHROMagar is nobel media for isolation & differentiation of Medically important *Candida* species.<sup>27</sup>

CHROMagar allow selective identification of *Candida* and on the basis of color reactions and colony morphology.

- *C. albicans* gave distinctive apple green color colonies.
- *C. galbrata* gave dark pink colonies with pale edges.
- *C. tropicalis* gave metallic blue colonies.
- *C. krusei* gave lavender color colonies but had velvety texture.



**Fig. 3:** Different type of organism give different color colonies in CHROMagar.

Each species also gave rise to a variety of colonies colors ranging from pink to green to blue of different colony characteristics later identity of all the isolates was confirmed with biochemical tests.<sup>28,29</sup>

Swab inoculated in SDA or CHROMagar but in case of tissue, they were stained with calcuoflower white stain (CFW) & Acridine orange (AO) stain. CFW is non specific fluorochrome stain that binds

to fungi while AO is metachromatin stain that selective for nucleic stain.<sup>30</sup>

Germ-tube test used in the rapid identification that revealed in 2-hr with 87.1% sensitive and 100% specific for the identification of *C. albicans*.<sup>31,32</sup>

Now days Polymerize chain reaction which is most rapid test for detection of any *Candida* spp. On the basis of gene difference & also able to differentiate the spp.. For *C. albicans*, single pair of primer SC1F & SC1R use that amplifies a 670 bp fragment of KER1 gene.<sup>33</sup> After isolation of genomic DNAs from *Candida* spp., species specific primers used for repetitive sequence for PCR amplification.<sup>34</sup>

On the basis of sequence data from ITS1 & ITS2 regions of reference strain from *Candida* genus which are available in EMBL/GeneBank databases, the species-specific primers, Calb, Cgla, Ckru, Cpar, Ctro, Clus, Cgni & Club were designed for specifically identify 8 clinical associated candidaspps. *C. albicans*, *C. Krusei*, *C. galbrata*, *C. parapsilosis*, *C. tropicalis*, *C. guilliermondii*, *C. lusitaniae* and *C. dubliniensis* respectively. These are yeast specific universal primers UNI1 & UNI2 used to amplify regions 1 (ITS1) & (ITS2) that are mostly associated with disease causing *Candida*.<sup>35</sup>

## Result

ERG11 is prevalent in all of genes which are responsible for emergence of antifungal resistance in *Candida* & spread through the mode of horizontal gene transfer with in case of hospital admitted patients that may also lead to community acquired infection because after discharge patients act as carriers in society.

## Conclusion

All the classes of antifungal are now resistance, due to the emergence of large no. of genes such as ABC1,SNQ1,TPO3,ERG3, ERG5, ERG6 or many more which provided the resistances to *Candida* in few decades that become big problem all around the world health sector. Ignorance of fungus infection with over use is also responsible for this anti fungal resistance because there have not action plane against the emerging resistance. There is lots of necessity of new age antifungal that help to counter the antifungal infection with new mode of action of drugs which plays important role. Early diagnosis also plays important role because with proper & correct antifungal drugs treatment can decreased the chance for development of antifungal cases. For this CHROMagar & multiplex PCR plays

important role in advanced diagnosis of early *Candida* infection.

## References

1. Stop neglecting fungi. *Nat. Microbiol.* 2017, 2, 17120. [CrossRef].
2. B. E. Jackson, K. R. Wilhelmus, and B. M. Mitchell, "Genetically regulated filamentation contributes to *Candida albicans* virulence during corneal infection," *Microbial Pathogenesis*, vol. 42, no. 2-3, pp. 88-93, 2007. [CrossRef].
3. Coevolution of Morphology and Virulence in *Candida* Species Delma S. Thompson, Patricia L. Carlisle, and David Kadosh [CrossRef].
4. *Candida* Infections, Causes, Targets, and Resistance Mechanisms: Traditional and Alternative Antifungal Agents, Claudia Spampinato<sup>1,2</sup> and Dario Leonardi<sup>3,4</sup> [CrossRef].
5. Hay, R. J. (1999). The management of superficial candidiasis. *Journal of the American Academy of Dermatology*, 40(6), S35-S42.
6. Messina, Fernando, and Ricardo Negroni. "Subcutaneous abscess as a single manifestation of candidiasis." *Medical Mycology* 1, no. 1 (2015): 6.
7. Badiie, Parisa, and Zahra Hashemizadeh. "Opportunistic invasive fungal infections: diagnosis & clinical management." *The Indian journal of medical research* 139, no. 2 (2014): 195.
8. Pappas, Peter G., Michail S. Lionakis, Maiken Cavling Arendrup, Luis Ostrosky-Zeichner, and Bart Jan Kullberg. "Invasive candidiasis." *Nature Reviews Disease Primers* 4, no. 1 (2018): 1-20.
9. Whaley, Sarah G., Elizabeth L. Berkow, Jeffrey M. Rybak, Andrew T. Nishimoto, Katherine S. Barker, and P. David Rogers. "Azole antifungal resistance in *Candida albicans* and emerging non-*albicans* *Candida* species." *Frontiers in microbiology* 7 (2017): 2173.
10. Nett, Jeniel E., and David R. Andes. "Antifungal agents: spectrum of activity, pharmacology, and clinical indications." *Infectious Disease Clinics* 30, no. 1 (2016): 51-83.
11. Ghannoum, Mahmoud A., and Louis B. Rice. "Antifungal agents: mode of action, mechanisms of resistance, and correlation of these mechanisms with bacterial resistance." *Clinical microbiology reviews* 12, no. 4 (1999): 501-517.
12. Kanafani, Zeina A., and John R. Perfect. "Resistance to antifungal agents: mechanisms and clinical impact." *Clinical infectious diseases* 46, no. 1 (2008): 120-128.
13. Birnbaum, Jay E. "Pharmacology of the allylamines." *Journal of the American Academy of Dermatology* 23, no. 4 (1990): 782-785.
14. Cowen, Leah E., Dominique Sanglard, Susan J. Howard, P. David Rogers, and David S. Perlin. "Mechanisms of antifungal drug resistance." *Cold Spring Harbor perspectives in medicine* 5, no. 7 (2015): a019752.
15. Hubbard, R. D., and S. Fidanze. "Therapeutic areas II: cancer, infectious diseases, inflammation and immunology and dermatology." *Comprehensive Medicinal Chemistry II* (2007).
16. Long, Sarah S., Charles G. Prober, and Marc Fischer. *Principles and practice of pediatric infectious diseases E-Book*. Elsevier Health Sciences, 2017.
17. Vanden Bossche H. Mechanisms of antifungal resistance. *Rev Iberoam Micol.* 1997 Jun;14(2):44-9. PMID: 16854169.
18. Sanglard, Dominique. "Emerging threats in antifungal-resistant fungal pathogens." *Frontiers in medicine* 3 (2016): 11.
19. Warnock, David W. "Antifungal drug susceptibility testing." *Current topics in medical mycology* (1989): 403-416.
20. Ksiezopolska, Ewa, and Toni Gabaldón. "Evolutionary emergence of drug resistance in *Candida* opportunistic pathogens." *Genes* 9, no. 9 (2018): 461.
21. Posteraro, Brunella, Riccardo Torelli, Antonietta Vella, Paolo Maria Leone, Giulia De Angelis, Elena De Carolis, Giulio Ventura, Maurizio Sanguinetti, and Massimo Fantoni. "Pan-echinocandin-resistant *Candida glabrata* bloodstream infection complicating COVID-19: A fatal case report." *Journal of Fungi* 6, no. 3 (2020): 163.
22. Zhu, Na, Dingyu Zhang, Wenling Wang, Xingwang Li, Bo Yang, Jingdong Song, Xiang Zhao et al. "A novel coronavirus from patients with pneumonia in China, 2019." *New England journal of medicine* (2020).
23. Chowdhary, Anuradha, Bansidhar Tarai, Ashutosh Singh, and Amit Sharma. "Multidrug-resistant *Candida auris* infections in critically ill coronavirus disease patients, India, April-July 2020." *Emerging Infectious Diseases* 26, no. 11 (2020): 2694.
24. Chowdhary, Anuradha, and Amit Sharma. "The lurking scourge of multidrug resistant *Candida auris* in times of COVID-19 pandemic." *Journal of Global Antimicrobial Resistance* 22 (2020): 175.
25. Arastehfar, Amir, Agostinho Carvalho, M. Hong Nguyen, Mohammad Taghi Hedayati, Mihai G. Netea, David S. Perlin, and Martin Hoenigl. "Covid-19-associated candidiasis (CAC): an underestimated complication in the absence of immunological predispositions?." *Journal of Fungi* 6, no. 4 (2020): 211.
26. Safavieh, Mohammadali, Chad Coarsey, Nwadiuto Esiobu, Adnan Memic, Jatin Mahesh Vyas, Hadi Shafiee, and Waseem Asghar. "Advances in *Candida* detection platforms for clinical and

- point-of-care applications." *Critical reviews in biotechnology* 37, no. 4 (2017): 441-458.
27. Odds, Frank C., and R. I. A. Bernaerts. "CHROMagar Candida, a new differential isolation medium for presumptive identification of clinically important Candida species." *Journal of clinical microbiology* 32, no. 8 (1994): 1923-1929.
  28. Pfaller, Michael A., Alasdair Houston, and S. Coffmann. "Application of CHROMagar Candida for rapid screening of clinical specimens for *Candida albicans*, *Candida tropicalis*, *Candida krusei*, and *Candida (Torulopsis) glabrata*." *Journal of clinical microbiology* 34, no. 1 (1996): 58-61.
  29. Madhavan, P., F. Jamal, P. P. Chong, and K. P. Ng. "Identification of local clinical *Candida* isolates using CHROMagar Candida TM as a primary identification method for various *Candida* species." *Tropical biomedicine* 28, no. 2 (2011): 269-274.
  30. Punjabi, Vishal, Shilpa Patel, Jigna Pathak, and Niharika Swain. "Comparative evaluation of staining efficacy of calcofluor white and acridine orange for detection of *Candida* species using fluorescence microscopy—A prospective microbiological study." *Journal of Oral and Maxillofacial Pathology: JOMFP* 24, no. 1 (2020): 81.
  31. Terrence Dolan, C., and Darlene M. Ihrke. "Further studies of the germ-tube test for *Candida albicans* identification." *American journal of clinical pathology* 55, no. 6 (1971): 733-734.
  32. Sheppard, Donald C., Marie-Claude Locas, Christiane Restieri, and Michel Laverdiere. "Utility of the germ tube test for direct identification of *Candida albicans* from positive blood culture bottles." *Journal of clinical microbiology* 46, no. 10 (2008): 3508-3509.
  33. Galán, Amparo, Verónica Veses, Amelia Murgui, Manuel Casanova, and José P. Martínez. "Rapid PCR-based test for identifying *Candida albicans* by using primers derived from the pH-regulated *KER1* gene." *FEMS yeast research* 6, no. 7 (2006): 1094-1100.
  34. Kanbe, Toshio, Keisuke Kurimoto, Hisao Hattori, Takako Iwata, and Akihiko Kikuchi. "Rapid identification of *Candida albicans* and its related species *Candida stellatoidea* and *Candida dubliniensis* by a single PCR amplification using primers specific for the repetitive sequence (RPS) of *Candida albicans*." *Journal of dermatological science* 40, no. 1 (2005): 43-50.
  35. Carvalho, Agostinho, S. Costa-De-Oliveira, M. L. Martins, C. Pina-Vaz, A. G. Rodrigues, Paula Ludovico, and F. Rodrigues. "Multiplex PCR identification of eight clinically relevant *Candida* species." *Medical mycology* 45, no. 7 (2007): 619-627.
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